

Research Article

The synthesis of dopaminergic radioligands labelled with tritium and iodine-125

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Abstract: The preparation of dopaminergic ligands radiolabelled with either tritium or iodine-125 has been an extremely important undertaking to advance understanding of this critical receptor class and the topic is reviewed from 1976 through 2006. Although not an exhaustive compilation of references, the cited examples highlight the many methods employed to prepare these useful reagents. Copyright © 2007 John Wiley & Sons, Ltd.

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Introduction

Dopamine plays a critical neurotransmitter role in numerous mammalian physiological functions. As the dominant catechol neurochemical, it exerts its exquisite multiple effects by means of transmembrane G protein-coupled receptors. Based on biochemical evidence, this receptor family has been further divided into subtypes referred to as the D1, D2, D3, D4 and D5 receptors and some of these may exist in both highand low-affinity states. However, even after decades of study, a complete understanding of the functional roles for all these individual receptor classes still appears incomplete. Abnormalities in the activity of dopamine receptors are thought to be in part responsible for such terrible diseases as schizophrenia, Parkinson's disease, attention deficit disorder and drug abuse among others. Therefore, the identification of potent dopaminergic agonists and antagonists has proven crucial to the discovery of safe and effective drugs in this area.¹ Clearly, the synthesis and characterization of radiolabelled dopaminergic ligands with tritium and iodine-125 have played a significant role in this important drug development process. This review will explore the successful techniques to label the structurally diverse collection of dopaminergic radioligands with tritium and iodine-125. It is not intended to be an exhaustive

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coverage of the voluminous literature in this field, but will showcase representative examples of synthetic strategy while spanning the momentous time period of 1976 to the end of 2006.

Tritiated dopaminergic radioligands

As a timely summary of the many methods to label compounds with tritium, the review by Saljoughian is recommended.² The methods related in this discussion for dopaminergic radioligands are essentially placed in the order of increasing specific activity outcome.

Tritium exchange reactions

Although rarely utilized because of the attendant low specific activity, tritium exchange reactions either with tritium gas or tritiated water and with or without catalyst have occasionally been used to prepare tritiated dopaminergic radioligands. Muccino and coworkers described the first tritium labelling of the antipsychotic haloperidol at very low specific activity using tritiated water³ and a similar procedure was used to label (*R*)-(–)-apomorphine and (*R*)-(–)-*N*-propylnor-apomorphine.⁴ Recently, the low specific activity exchange tritiation of dopamine itself was reported as well.⁵



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Catalytic tritium dehalogenation

Because many dopaminergic compounds contain electron-rich aromatic rings which can be readily halogemethods. catalvtic nated bv various tritium dehalogenation has remained a useful method of choice for tritiation of these ligands at reasonably high specific activity. We were surprised to learn that the preparation of tribromodopamine (1) had never been described, but it proved to be easily synthesized and a very convenient precursor to afford [ring-³H] dopamine at 44 Ci/mmol by catalytic (5% Pd/C) tritium debromination.⁶ In this same report we described the tritiation of several fluorinated congeners as well. Using a similar approach with precursor **2**, we also obtained $[8.9-^{3}H]$ (R)-(–)-apomorphine.⁷ We employed a catalytic tritium dehalogenation scheme for the preparation of the irreversible agonist [8,9-³H] (R)-(-)-N-chloroethylnorapomorphine⁸ and $[5,8^{-3}H]$ (+/-)-6,7-ADTN⁹ too. In all of these cases, proton-decoupled tritium NMR was utilized to characterize the products.



The 2,3,4,5-tetrahydro-1H-3-benzazepine class of compounds, decorated with a variety of diverse functionality, has been a rich source of valuable dopaminergic ligands, especially D1 receptor antagonists. As a member of this class, (+/-)-SCH 23390 contains both a chlorine and hydroxyl group in the 7 and 8 positions, respectively, and was the subject of a tritiation study.¹⁰ Hoping to exploit the presence of the large cyclic tertiary amine, a Leonard oxidation¹¹ of it to the enamine was first attempted. This approach had been used in the past with great success in the tritiation of such morphinan ring systems as $[15, 16^{-3}H]$ buprenorphine.¹² However, attempts at using mercuric oxide to create an enamine analogue of (+/-)-SCH 23390 inexplicably failed and the authors turned to a catalytic tritium dehalogenation approach. Bromination of (+/-)-SCH 23390 afforded the monobromo analogue in modest yield with halogenation occurring as expected only ortho to the phenol. Catalytic tritium dehalogenation for a short time using 5% Pd/C was selective enough to effect exclusive debromination and provide the desired product in modest yield

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and specific activity. In another report, two other compounds structurally related to (+/-)-SCH 23390 were also successfully tritiated.¹³ (R.S)-SKF 82526 containing multiple hydroxyl groups was solely monobrominated in the 4-hydroxyphenyl ring and subsequently catalytically tritium debrominated to give [³H] (R,S)-SKF 82526 at 15-23 Ci/mmol. In an attempt to elevate its specific activity and enhance receptorbinding performance, the R enantiomer alone of this compound was also diiodinated on the 4-hydroxyphenyl ring with ICl and tritiated in analogous manner to obtain [³H] (*R*)-SKF 82526. However, both forms of [³H] SKF 82526 were found to be rather unstable and storage solvent studies were conducted to minimize decomposition. The potential racemization of its benzylic chiral center during the tritiation was arguably ruled out by demonstrating that the chiral integrity was maintained during a parallel catalytic hydrogen dehalogenation. In the same paper an identical strategy was also used to prepare structurally related [³H] SKF 38393.

The benzamide class of drugs has been another productive area to identify useful dopaminergic ligands. Sulpiride, a member of this group, was first successfully tritiated by a catalytic tritium debromination method at moderate specific activity.14 Interestconventional bromination conditions for ingly. sulpiride failed, but it could be successfully brominated with a mixture of hydrogen bromide and hydrogen peroxide. A similar method was also employed by these same authors to prepare the related drug [³H] sultopride. Other benzamide dopaminergics that have been radiolabelled by catalytic tritium dehalogenation include NCQ 115¹⁵ and a selective D4 receptor agonist by Abbott radiochemists.¹⁶ In this last example, catalytic tritium dehalogenation of a tetrabromo precursor afforded the desired product at a high specific activity of 88 Ci/mmol.

Bromine and iodine have not been the only halogens to serve as valuable precursor functionality for catalytic tritium dehalogenation. Under certain circumstances even chlorine has been employed. For instance, the dopamine reuptake inhibitor mazindol has been tritiated by catalytic tritium dechlorination.¹⁷ Also, we prepared and characterized [benzoyl-3,4-³H] cocaine (3), utilizing a convenient dichlorophenyl precursor. Its proton-decoupled tritium NMR demonstrating exclusive aromatic tritium labelling is shown in Figure 1. Thurkauf reported the synthesis of the selective D4 receptor radioligand [³H] NGD 94-1 (4) at a specific activity of 57 Ci/mmol using a multiple bromo dichloro iodo precursor.¹⁸ The goal of this project was the development of a radioligand that could be used to explore the role of the D4 receptor in



Figure 1 Proton decoupled tritium NMR (CD₃OD) of 3.

schizophrenia and other neurological diseases but with greater D4 receptor selectivity than [³H] clozapine.



The strategy of catalytic tritium dehalogenation raises the intriguing issue of possible chemoselectivity in the process; namely, selective tritium dehalogenation without reduction of other accompanying functionality such as olefins, nitro and cyano groups or other halogens. The selective catalytic tritiation of an aryl bromide or iodide precursor also containing an aryl chlorine has already been described above for 5% Pd/ C¹⁰ and Lindlar catalyst (Pd/CaCO₃).¹³ Also, the catalytic tritium polydebromination of a nitrile containing precursor has been reported.¹⁶ Pfizer investigators also utilized Rosenmund catalyst (Pd/BaSO₄) to accomplish a selective catalytic tritium debromination in the presence of an aryl chlorine at 40–50 psi.¹⁹ Demonstrating the chemoselectivity of a catalytic tritium debromination in the presence of an olefin, we also employed Lindlar catalyst on precursor 5 to obtain $[2^{-3}H]$ (–)-lisuride (6), a D2 receptor agonist.²⁰ Again,

proton-decoupled tritium NMR provided conclusive evidence of the exclusive 2-position tritium labelling of **6**. Other recently published reports²¹⁻²³ using sodium borotritide in conjunction with certain transition metal catalysts should also provide a valuable tool to effect such chemoselective tritium dehalogenations as they are required in this and other radioligand areas.



Catalytic tritiation of unsaturated positions

The catalytic tritiation of various unsaturated precursors has also been an effective method to prepare many dopaminergic radioligands. Several benzamide class dopaminergics were tritiated at higher specific activity by means of a general and efficient approach as reported in publications from the Astra Lakemedel laboratory.^{24–26} Taking advantage of the fact that a chiral pyrrolidine moiety is prevalent in many compounds of this group, these



Figure 2 Proton decoupled tritium NMR (CD₃OD) of 8.

workers prepared a common precursor, (S)-N-acetyl-3, 4-dehydroprolinamide and catalytically tritiated it to the corresponding [3,4-pyrrolidine-³H] (S)-2-(aminomethyl)-1-ethylpyrrolidine. This intermediate was then condensed with various functionalized benzoic acid derivatives to obtain an ensemble of interesting products. In this way, tritiated versions of such compounds as raclopride, eticlopride and remoxipride with specific activities as high as 58 Ci/mmol were prepared. In the case of $[3,4-pyrrolidine-{}^{3}H]$ remoxipride, the location of tritium labelling was clearly elucidated by proton-decoupled tritium NMR and the observed tritium-tritium coupling constant of 10.3 Hz was consistent with the assigned doubly labelled structure. Also, the integrity of the chiral center was inferred by demonstrating the lack of racemization in a corresponding catalytic hydrogenation.

A tritiated photoaffinity analogue of the benzamide dopaminergic sulpride was also prepared by a related but slightly different strategy; namely, the conversion of an allyl group to a tritiated propyl group.²⁷ The use of an appended allyl group has been a convenient entry to other valuable tritiated dopaminergic radioligands. We employed this method both in the preparation of $[N-\text{propyl}-^{3}\text{H}]$ (-)-*N*-propylnorapomorphine (7) 28 as well as [*N*-propyl- 3 H] quinpirole (**8**), a radioligand with high affinity for the D2 receptor subclass.²⁹ For product 7, proton-decoupled tritium NMR also corroborated the exclusive tritium labelling of the propyl group. A very similar tritium NMR for radioligand 8 is shown in Figure 2. This same allyl tritiation methodology was further utilized in the labelling of D2 agonists U-91356 and U-86170,30,31 the latter containing two tritiated propyl groups. We were also able to introduce two tritiated N-propyl pendants in the synthesis and characterization of the selective D3 receptor agonist $[^{3}H]$ (+/-)-7-hydroxy DPAT.³² In our case the concomitant tritiation of a cyclic enamine conferred even higher specific activity on the product. Other tritiations that have exercised this strategy were the radiolabelling of the dopamine agonist pergolide by Wheeler and co-workers at Eli Lilly³³ and the D2 receptor agonist (+)-PHNO by



Figure 3 Proton decoupled tritium NMR (CD₃OD) of [*N*-propyl-³H] (+)-3-PPP.

Seeman and colleagues at the University of Toronto.³⁴ Besides the allyl group, the propargyl functionality has also facilitated the installation of tritiated propyl groups at even higher specific activity. This methodology was used to tritiate the D3 receptor agonist PD 128907³⁵ and the D2 receptor agonist N-0437 as a racemic mixture.³⁶ We later employed a chiral Npropargyl analogue of N-0923 to prepare [N-propyl-³H] N-0923, the (-) enantiomer of N-0437 with the most potent D2 receptor activity.37 Again, its proton-decoupled tritium NMR documented the strictly propyl radiolabelling. Although the dopamine autoreceptor agonist (-)-3-PPP had been previously tritiated at modest specific activity by means of a ring olefin analogue.³⁸ we were able to prepare its other enantiomer [N-propyl-³H] (+)-3-PPP, a sigma receptor radioligand, at greater than 80 Ci/mmol using an N-propargyl precursor. Its proton-decoupled tritium NMR is shown in Figure 3. This same strategy in principle could be employed to tritiate (-)-3-PPP at high specific activity.



Because of its connection to the growing problem of cocaine abuse, the dopamine reuptake (or transporter)

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complex and compounds interacting with it have been very actively investigated. Such radioligands have been prepared by catalytic tritiation of olefin precursors and these include BTCP,³⁹ fourphit,⁴⁰ NNC 12-0781,⁴¹ as well as our tritium labelling of O-972.⁴² Workers at the NIMH also adopted this labelling strategy to prepare one of the most potent dopamine reuptake blockers, [³H] GBR-12935 (**9**),⁴³ as well as a tritiated azido photoaffinity analogue (**10**) of it.⁴⁴ These latter radioligands have been exceptionally valuable since they do not bind to the norepinephrine or serotonin transporter complexes like other structurally related compounds.



Installation of the $C^{3}H_{3}$ group

Many dopaminergic radioligands, including some of those already mentioned, have pendant methyl groups incorporated in their structures, often attached to nitrogen or as methyl esters and ethers. This convenient structural circumstance has presented an outstanding opportunity to prepare tritiated methyl versions of these compounds by various means. Often these groups have been added by an electrophilic reaction using the reagent tritiated methyl iodide. However, Pounds has recently introduced a valuable alternative reagent, [³H] methyl nosylate,^{45,46} which has demonstrated even more advantages for such labelling, especially on a small scale where stoichiometry is often crucial.

The benzazepine SCH 23390 has been labelled as an $N-C^{3}H_{3}$ analogue with tritiated methyl iodide.⁴⁷ Also, Mailman and colleagues later improved upon the specific activity of this radioligand by first catalytic tritium debromination of a desmethyl precursor followed by N-methylation with tritiated methyl iodide to achieve an enhanced specific activity of 93.8 Ci/ mmol.⁴⁸ Other dopaminergics labelled as N-C³H₃ derivatives include clozapine,⁴⁹ U-101958,⁵⁰ WIN 35,065-2,⁵¹ and our labelling of WIN 35,428 for study of the dopamine reuptake site.^{52,53} There have also been several reports of dopaminergics radiolabelled with $O-C^{3}H_{3}$ groups and these include the methyl ester labelling of dopamine transport ligands [methyl ester-³H] beta-CIT and fluoro analogues,⁵⁴ [methyl ester-³H] LTB-999 (11)⁵⁵ the benzamide [methoxy-³H] raclopride⁵⁶ along with our own synthesis of [meth $oxy^{-3}H$] (–)-sulpiride (**12**).



Iodine-125-labelled dopaminergic radioligands

Over the past 30 years, along with their tritiated counterparts, numerous dopaminergic radioligands labelled with iodine-125 have also made a significant contribution in this area. Several useful reviews of the various methods to introduce radioiodine into organic molecules have been published^{57,58} along with the comprehensive paper by Bolton on all radiohalogenation techniques with a significant section on radioiodination.59 Because of the relatively smaller molecular size of dopaminergic compounds, bulky prosthetic groups like the classic Bolton-Hunter reagent are rarely used and it is often a single ¹²⁵I itself that is introduced into the parent structure. However, having said that, it is rather surprising that the attachment of such a large atom like iodine to an established dopaminergic compound scaffold could still result in the creation of a useful and effective radioligand.

Electrophilic iodine-125 radiolabelling

There are numerous instances of the direct radioiodination of dopaminergic ligands with iodine-125, employing some oxidation process on [125I] sodium iodide. Early work at then SmithKline & French laboratories in the preparation of [¹²⁵I] iodospiroperidol is an excellent case study of the thoughtful planning and exacting experimental design that often accompanies such radiolabelling.⁶⁰ Encouraged by the report that [⁷⁷Br] *p*-bromospiroperidol was successfully synthesized at Argonne National Laboratory and utilized for in vivo imaging with its distribution much like that of [³H] spiroperidol itself,^{61–63} Landvatter explored the preparation of [¹²⁵I] iodospiroperidol for both *in vivo* as well as in vitro applications. He found that the oxidation conditions required for its radiolabelling were very pH sensitive, requiring careful control of the buffer environment. When phosphate buffer was substituted for potassium acetate buffer, no reaction occurred and pH ranges that varied from the optimum 3.85 gave sharply reduced yields. When the oxidant chloramine-T (CAT) was substituted for hydrogen peroxide, the side product *p*-chlorospiroperidol was formed. The paper further described an efficient single HPLC purification procedure to afford pure product at high specific activity.

Conveniently, the compound SCH 23982, a member of the benzazepine class of dopaminergics, already contains an iodine atom. It had previously been radiolabelled with $^{123}I^{64,65}$ and $^{125}I^{66}$ for use as a high-affinity D1 receptor antagonist. However, Mailman's group further improved the radioiodination process by a detailed and robust experimental protocol with increased yields and a simpler purification method.⁶⁷ They also unequivocally characterized the radioiodinated product by demonstrating that its diode array detected UV spectrum was completely superimposable on that of authentic unlabelled SCH 23982. As in the previous case of spiroperidiol, when the authors employed CAT as the oxidant for the [¹²⁵I] sodium iodide, they noted that a side product in the synthesis was the unlabelled chloro isomer SCH 23390. In this same receptor area and compound class there have been several reports of radioiodinated azido photoaffinity ligands for the D1 receptor emerging from the collaboration of the Neumeyer and Seeman groups along with our technical assistance.^{68,69}

Other examples of dopaminergic ligands that have been directly radioiodinated are BZM,^{70,71} NCQ 298⁷²

and both alpha and beta-CIT.73 As mentioned, there have been many oxidants employed in direct radioiodination and in the case of BZM the authors examined a number of them including CAT, hydrogen peroxide, sodium persulphate, metachloroperoxybenzoic acid and peracetic acid. The latter was found to be a superior oxidant with a higher vield, reduced reaction time and enhanced radiochemical purity.⁷¹ These authors found this method advantageous for iodine-123 labelling as well. Other examples using direct radioiodination in this way include the irreversible compounds for the dopamine transporter complex, photoaffinity ligands [¹²⁵I] RTI- 82^{74} and $[^{125}I]$ MFZ-2-24 (13) as well as the alkylating agent [125I] MFZ-3-37 (14).75 In the case of these latter two radioligands, a common aniline derivative served as the branch point intermediate which was first radioiodinated and then converted into either the azide or isothiocyanate functionalized product. Very recently these same NIDA workers also reported the radioiodination of another cocaine analogue containing a biphenvl azide at the 3-beta position.⁷⁶ The strategy behind this effort was to vary the position of the azido group on the cocaine structure scaffold, possibly allowing radioligand attachment points on the dopamine transporter to be identified as an aid in novel drug design.



The previous literature citations all dealt with direct radioiodination of a particular dopaminergic ligand substrate and, more often than not, an aromatic group. Another popular strategy used in electrophilic radioiodination is destannylation; that is, the activation of an aromatic group as an organotin derivative prior to radiolabelling. Organotin compounds are easily prepared by a number of efficient organometallic reactions and radioiododestannylation has proved to be a robust and reliable synthetic method.⁷⁷ Several benzazepine D1 receptor ligands were radioiodinated in this way by the Kung group^{78,79} as well as the dopamine transporter ligands beta-CIT,54 RTI-12180 and structurally related RTI-229.81 Also, several benzamide class dopaminergics have been radioiodinated using this versatile methodology as well. $^{82-86}$

Although early in this discussion it was mentioned that the relatively small size of dopaminergic substrates usually ruled out the use of most added bulky radiolabelled appendages like the Bolton-Hunter reagent, one prosthetic group that has been successfully employed for radioiodinations is the vinylstannyl group. This labelling option has been exceptionally valuable in preparing target compounds that required radioiodination in positions other than aromatic rings. In large part Hanson and colleagues working jointly at Northeastern University and the Harvard Medical School pioneered the early introduction of the vinylstannyl group for radioiodination via nucleophilic additions to carbonyl compounds.87,88 In complimentary fashion, Lever and co-workers at Johns Hopkins later introduced electrophilic cis and trans vinylstannyl tosylate derivatives followed by radioiodination to label spiroperidol analogues 15 and 16, respectively.^{89,90} These radioiodinations occurred under mild conditions and with retention of the stannyl regio and steric configurations. The authors also examined whether their radioiodinated products might be contaminated by any unlabelled side products. For that reason both E and Z chloro analogues of compounds 15 and 16 were separately prepared (since they could have resulted from a CAT promoted chlorodestannylation), but these side products were not observed by HPLC. However, the allyl derivative (R1 = R2 = H) resulting from the protodestannylation was also synthesized and found to be present in the crude radioiodinated mixture. Fortunately, this allyl side product was well resolved from both compounds 15 and 16 and could be easily removed by reverse-phase HPLC purification. This electrophilic installation of vinylstannyl precursors has also been successfully employed in the radioiodination of several dopamine transporter ligands.^{91,92} Finally, another successful method to introduce the vinylstannyl group has been a radical catalyzed addition of a tin reagent to a preexisting alkyne group as exploited by the Kung laboratory.93-96



16 R1 = H, R2 = 125

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Nucleophilic iodine-125 radiolabelling

A second general method that has been useful to radioiodinate dopaminergic compounds has been by nucleophilic means and usually employing [125 I] sodium iodide. In this way the Nakatsuka group at Sumitomo utilized a radioiodine exchange to produce [2^{-125} I] 2-iodospiroperidol and the radioligand showed promise for both *in vitro* and *in vivo* dopamine receptor study.^{97–101} Also, several benzamide dopaminergics^{102,103} as well as mazindol¹⁰⁴ were radioiodinated in like fashion. Sulpride was radioiodinated by [125 I] sodium iodide displacement of a diazotized aniline precursor.¹⁰⁵ Finally, [125 I] iodoethylspiroperidol was prepared by [125 I] sodium iodide displacement of a tosylate group.^{106,107}

Conclusion

Although dopaminergic ligands have been tagged with other stable and radioactive isotopes, this review has focused only on those radiolabelled with tritium and iodine-125 during the past 30 years, a significant time period for dopaminergic receptor binding assay development. Many of the radioligands discussed here each have scores of published papers on their use and application in biological assays, but the citations included in this review were selected for their relevance in highlighting the synthetic strategy for this structurally diverse ensemble of compounds. It is hoped that this coverage will not only provide an interesting historical perspective but also a useful guide for future radiosynthetic progress in this important receptor area.

REFERENCES

- 1. Zhang A, Kan Y, Li F. *Expert Opin Ther Patents* 2006; **16**: 587–630.
- 2. Saljoughian M. Synthesis 2002; 1781–1801.
- Muccino RR, Serico L. J Label Compd Radiopharm 1978; 15: 523–527.
- Soine WH, Salgo P, Smith RV. J Label Compd Radiopharm 1979; 16: 597–601.
- Pajak M, Kanska M. J Label Compd Radiopharm 2006; 49: 1061–1067.
- Filer CN, Orphanos D, Seguin RJ. Synth Commun 2006; 36: 975–978.
- Filer CN, Ahern DG. J Org Chem 1980; 45: 3918–3919.
- Guan J-H, Neumeyer JL, Filer CN, Ahern DG, Lilly L, Watanabe M, Grigoriadis D, Seeman P. J Med Chem 1984; 27: 806–810.

- Orphanos D, Filer CN. J Radioanal Nucl Chem 2002; 254: 223–224.
- Wyrick SD, Mailman RB. J Label Compd Radiopharm 1985; 22: 189–195.
- 11. Leonard NJ, Hay AS, Fulmer RW, Gash VW. J Am Chem Soc 1955; **77**: 439–444.
- 12. Otvos F, Hosztafi S, Simon C, Toth G. J Label Compd Radiopharm 1995; **36**: 79–83.
- Landvatter SW, Blackburn DW, Villani AJ, Bosch GK. J Label Compd Radiopharm 1987; 24: 145–155.
- Pri-Bar I, Buchman O. J Label Compd Radiopharm 1983; 20: 1153–1158.
- Hall H, Hogberg T, Halldin C, Bengtsson S, Wedel I. Eur J Pharmacol 1991; 201: 1–10.
- Matulenko MA, Surber B, Fan L, Kolasa T, Nakane M, Terranova MA, Uchic ME, Miller LN, Chang R, Donnelly-Roberts DL, Namovic MT, Moreland RB, Brioni JD, Stewart AO. *Bioorg Med Chem Lett* 2004; **14**: 5095–5098.
- 17. Dugger HA, Tabot KC. *J Label Compd Radiopharm* 1976; **12**: 429–436.
- Thurkauf A. J Label Compd Radiopharm 1997;
 39: 123–128.
- Howard HR, Shenk KD, Smolarek TA, Marx MH, Windels JH, Roth RW. J Label Compd Radiopharm 1994; 39: 117–125.
- Filer CN, Hainley C, Nugent RP. J Radioanal Nucl Chem 2006; 267: 345–348.
- 21. Nagasaki T, Sakai K, Segawa M, Katsuyama Y, Haga N, Koike M, Kawada K, Takechi S. *J Label Compd Radiopharm* 2001; **44**: 993–1004.
- Tang YS, Lui W, Chaudhary A, Melillo DG, Dean DC. Synthesis and Applications of Isotopically Labelled Compounds, vol. 8. Wiley: Chichester, 2004; 71–74.
- 23. Chaudhary A, Tang YS, Dean DC, Ashton W, Melillo DG. Synthesis and Applications of Isotopically Labelled Compounds, vol. 8. Wiley: Chichester, 2004; 425–428.
- Gawell L, Hall H, Kohler C. J Label Compd Radiopharm 1985; 22: 1033–1043.
- 25. Kohler C, Hall H, Ogren S-O, Gawell L. *Biochem Pharmacol* 1985; **34**: 2251–2259.
- Hall H, Kohler C, Gawell L. Eur J Pharmacol 1985; 111: 191–199.
- Rognan D, Mann A, Hamdi P, Wermuth CG, Sokoloff P, Schwartz JC, Roy J, Morgat JL. J Label Compd Radiopharm 1987; 24: 1361–1372.
- Filer CN, Ahern DG, Granchelli FE, Neumeyer JL, Law S-J. J Org Chem 1980; 45: 3465–3467.
- Seeman P, Schaus JM. Eur J Pharmacol 1991;
 203: 105–109.

- Moon MW, Hsi RSP. J Label Compd Radiopharm 1992; 31: 933–943.
- Moon MW, Morris JK, Heier RF, Hsi RSP, Manis MO, Royer ME, Walters RR, Lawson CF, Smith MW, Lahti RA, Piercey MF, Sethy VH. Drug Des Discov 1993; 9: 313–322.
- Pounds S, Seguin RJ, Filer CN. Appl Radiat Isot 2005; 62: 49–53.
- Wheeler WJ, Kau DLK, Bach NJ. J Label Compd Radiopharm 1990; 28: 273–295.
- Seeman P, Ulpian C, Larsen RD, Anderson PS. Synapse 1993; 14: 254–262.
- Akunne HC, Towers P, Ellis GJ, Dijkstra D, Wikstrom H, Heffner TG, Wise LD, Pugsley TA. *Life Sci* 1995; **57**: 1401–1410.
- Van der Weide J, De Vries JB, Tepper PG, Horn AS. Eur J Pharmacol 1987; 134: 211–219.
- 37. Laseter AG, Seguin RJ, Filer CN. *J Radioanal Nucl Chem* 2005; **264**: 723–725.
- Thorberg S-O, Johansson L, Gawell L, Sahlberg C. J Label Compd Radiopharm 1986; 23: 927–934.
- de Costa BR. J Label Compd Radiopharm 1991;
 29: 165–173.
- Linders JTM, de Costa BR, Grayson NA, Rice KC. J Label Compd Radiopharm 1992; 31: 671–683.
- Muller L, Hall H, Halldin C, Farde L, Hohlweg R, Suzdak PD, Nielsen EB, Foged C. *Nucl Med Biol* 1995; **22**: 711–717.
- 42. Dutta AK, Reith MEA, Madras BK. Synapse 2001;39: 175–181.
- Berger P, Janowsky A, Vocci F, Skolnick P, Schweri MM, Paul SM. Eur J Pharmacol 1985; 107: 289–290.
- 44. Thurkauf A, de Costa BR, Berger P, Paul SM, Rice KE. J Label Compd Radiopharm 1991; **29**: 125–129.
- 45. Pounds S. Synthesis and Applications of Isotopically Labelled Compounds, vol. 8. Wiley: Chichester, 2004; 469–472.
- Pounds S. United States Provisional Patent Serial No. 60/320,238 filed 30 May 2003.
- 47. Billard W, Ruperto V, Crosby G, Iorio LC, Barnett A. *Life Sci* 1984; **35**: 1885–1893.
- 48. Wyrick SD, McDougald DL, Mailman RB. J Label Compd Radiopharm 1986; **23**: 685–692.
- Sunay UB, Talbot KC, Galullo V. J Label Compd Radiopharm 1992; 31: 1041–1047.
- Helmeste DM, Shioiri T, Mitsuhashi M, Tang SW. Eur J Pharmacol 1999; 370: 205–209.
- Naseree TM, Abraham P, Kepler JA, Carroll FI, Lewin AH, Kuhar MJ. J Label Compd Radiopharm 1990; 28: 1011–1016.

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- 52. Lewin AH, Gao Y, Abraham P, Boja JW, Kuhar MJ, Carroll FI. *J Med Chem* 1992; **35**: 135–140.
- 53. Abraham P, Pitner JB, Lewin AH, Boja JW, Kuhar MJ, Carroll FI. *J Med Chem* 1992; **35**: 141–144.
- Swahn C-G, Halldin C, Gunther I, Patt J, Ametamey S. J Label Compd Radiopharm 1996; 38: 675–685.
- 55. Dolle F, Emond P, Mavel S, Demphel S, Hinnen F, Mincheva Z, Saba W, Valette H, Chalon S, Halldin C, Helfenbein J, Legaillard J, Madelmont J-C, Deloye J-B, Bottlaender M, Guilloteau D. *Bioorg Med Chem* 2006; **14**: 1115–1125.
- Ehrin E, Gawell L, Hogberg T, de Paulis T, Strom P. J Label Compd Radiopharm 1987; 24: 931–940.
- Seevers RH, Counsell RE. Chem Rev 1982; 82: 575–590.
- 58. Baldwin RM. Appl Radiat Isot 1986; **37**: 817–821.
- Bolton R. J Label Compd Radiopharm 2002; 45: 485–528.
- Landvatter SW. J Label Compd Radiopharm 1985;
 22: 273–278.
- Kulmala HK, Huang CC, Dinerstein RJ, Friedman AM. Life Sci 1981; 28: 1911–1916.
- Friedman AM, Huang CC, Kulmala HK, Dinerstein RJ, Navone J, Brunsden B, Gawlas D, Cooper M. Int J Nucl Med Biol 1982; 9: 57–61.
- DeJesus OT, Friedman AM, Prasad A, Revenaugh JR. J Label Compd Radiopharm 1983; 20: 745–756.
- Moerlein SM, Parkinson D, Welch MJ. Appl Radiat Isot 1990; 41: 381–385.
- Beer HF, Lin S, Novak-Hofer I, Blaeuenstein P, Schubiger PA. Appl Radiat Isot 1992; 43: 781–787.
- Kung HF, Billings JJ, Guo YZ, Blau M, Ackerhalt R. Nucl Med Biol 1988; 15: 187–193.
- Lawler CP, Gilmore JH, Mooney DH, Mayleben MA, Atashi JR, Mileson BE, Wyrick SD, Mailman RB. J Neurochem Methods 1993; 49: 141–153.
- Baindur N, Neumeyer JL, Niznik HB, Bzowej NH, Jarvie KR, Seeman P, Garlick RK, Miller JJ. *J Med Chem* 1988; **31**: 2069–2071.
- Neumeyer JL, Baindur N, Yuan J, Booth G, Seeman P, Niznik HB. J Med Chem 1990; 33: 521–526.
- Kung HF, Kasliwal R, Pan S, Kung M-P, Mach RH, Guo Y-Z. J Med Chem 1988; 31: 1039–1043.
- Kung M-P, Kung HF. J Label Compd Radiopharm 1989; 27: 691–700.
- Hogberg T, Strom P, Hall H, Kohler C, Halldin C, Farde L. Acta Pharm Nord 1990; 1: 53–60.
- Muller L, Halldin C, Swahn C-G, Foged C. J Label Compd Radiopharm 1994; 34: 1031–1040.

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- Lever JR, Carrol FI, Patel A, Abraham P, Boja J, Lewin A, Lew R. J Label Compd Radiopharm 1993;
 33: 1131–1137.
- Lever JR, Zou M-F, Parnas ML, Duval RA, Wirtz SE, Justice JB, Vaughan RA, Newman AH. *Bioconjug Chem* 2005; 16: 644–649.
- Newman AH, Cha JH, Cao J, Kopajtic T, Katz JL, Parnas ML, Vaughan RA, Lever JR. J Med Chem 2006; 49: 6621–6625.
- Kabalka GW, Varma RS. Tetrahedron 1989; 45: 6601–6621.
- Chumpradit S, Kung HF, Billings JJ, Kung M-P, Pan S. J Med Chem 1989; **32**: 1431–1435.
- Chumpradit S, Kung M-P, Billings JJ, Kung HF. J Med Chem 1991; 34: 877–883.
- Lever JR, Scheffel U, Stathis M, Seltzman HH, Wyrick CD, Abraham P, Parham K, Thomas BF, Boja JW, Kuhar MJ, Carroll FI. *Nucl Med Biol* 1996; 23: 277–284.
- Zhong D, Kotian P, Wyrick CD, Seltzman HH, Kepler JA, Boja JW, Kuhar MJ, Carroll FI. J Label Compd Radiopharm 1999; 42: 281–286.
- de Paulis T, Janowsky A, Kessler RM, Clanton JA, Smith HE. J Med Chem 1988; 31: 2027–2033.
- Murphy RA, Kung HF, Kung M-P, Billings JJ. J Med Chem 1990; 33: 171–178.
- 84. de Paulis T, Smith HE. Synth Commun 1991; **21**: 1091–1095.
- Clanton JA, de Paulis T, Schmidt DE, Ansari MS, Manning RG, Baldwin RM, Kessler RM. J Label Compd Radiopharm 1991; 29: 745–751.
- Farouk N, Motaleb MA, Shweeta HA, Farah K, Kolaly MT. J Radioanal Nucl Chem 2005; 266: 405–410.
- Hanson RN, Seitz DE. J Label Compd Radiopharm 1982; 19: 1585–1586.
- 88. Hanson RN, Seitz DE, Botarro JC. J Nucl Med 1982; **23**: 431–436.
- Musachio JL, Lever JR. Tetrahedron Lett 1989;
 30: 3613–3616.
- 90. Musachio JL, Lever JR. *Bioconjug Chem* 1992; **3**: 167–175.
- Elmaleh DR, Fischman AJ, Shoup TM, Byon C, Hanson RN, Liang AY, Meltzer PC, Madras BK. J Nucl Med 1996; 37: 1197–1202.

- 92. Edmond P, Boazi M, Duchene A, Chalon S, Besnard JC, Guilloteau D, Frangin Y. J Label Compd Radiopharm 1997; 39: 757–772.
- 93. Canney DJ, Guo Y-Z, Kung M-P, Kung HF. J Label Compd Radiopharm 1993; **33**: 355–368.
- 94. Chumpradit S, Kung M-P, Kung HF. J Med Chem 1993; **36**: 4308–4312.
- Goodman MM, Kung M-P, Kabalka GW, Kung HF, Switzer R. J Med Chem 1994; 37: 1535–1542.
- 96. Chumpradit S, Kung M-P, Vessotskie J, Kung HF. *J Label Compd Radiopharm* 1995; **36**: 1051–1062.
- 97. Nakatsuka I, Shimizu H, Shono F, Yoshitake A, Saji H, Tokui T, Kuge U, Yokoyama A, Torizuka K. Synthesis and Applications of Isotopically Labelled Compounds. vol. 2. Wiley: Chichester, 1985; 153–154.
- Nakatsuka I, Saji H, Shiba K, Shimizu H, Okuno M, Yoshitake A, Yokoyama A. *Life Sci* 1987; **41**: 1989–1997.
- 99. Saji H, Nakatsuka I, Shiba K, Tokui T, Horiuchi K, Yoshitake A, Torizuka K, Yokoyama A. *Life Sci* 1987; **41**: 1999–2006.
- 100. Saji H, Shiba K, Saiga A, Tokui T, Nakatsuka I, Okuno M, Yoshitake A, Yokoyama A. J Label Compd Radiopharm 1989; 26: 95–97.
- Nakatsuka I, Shiba K, Saji H, Yoshitake A, Yokoyama A. J Label Compd Radiopharm 1993;
 33: 105–118.
- 102. Neumeyer JL, Guan J-H, Niznik HB, Dumbrille-Ross A, Seeman P, Padmanabhan S, Elmaleh DR. *J Med Chem* 1985; 28: 405–407.
- 103. Saji H, Tanahashi K, Kinoshita T, Iida Y, Magata Y, Yokoyama A. Nucl Med Biol 1996; 23: 121–127.
- 104. Galinier E, Ombetta JE, Frangin Y, Mertens J, Besnard JC, Guilloteau D. J Label Compd Radiopharm 1994; 34: 487–497.
- 105. Sales N, Martres MP, Bouthenet ML, Schwartz JC. Neuroscience 1989; 28: 673–700.
- 106. Frangin Y, Caillet M, Chalon S, Huguet F, Foulon C, Desplanches G, Baulieu J-L, Besnard J-C, Guilloteau D. J Label Compd Radiopharm 1990; 28: 1363–1371.
- 107. Chalon S, Frangin Y, Guilloteau D, Caillet M, Guimbal C, Schmitt M-H, Desplanches G, Baulieu J-L, Besnard J-C. *Nucl Med Biol* 1990; 17: 389–395.